PHOTOOXIDATION OF OLEANOLIC ACID: FORMATION OF 11a, 12a-EPOXY- AND 12a-HYDROXY-OLEANOLIC LACTONES

Isao Kitagawa, Kiyoshi Kitazawa, and Itiro Yosioka Faculty of Pharmaceutical Sciences, Osaka University

> Toyonaka, Osaka, Japan (Received in Japan 9 October 1967)

In 1965, Corey and his co-workers reported¹⁾ an interesting paper on the oxidative rearrangement of pentacyclic triterpenes which involved a novel skeletal rearrangement of thermodynamically more favored β-amyrin (I) towards less stable taraxerene derivative (II) by way of photochemical oxidative procedure in the acidic medium. They proposed an intermediate (III) in connection with the independent synthesis of the same product (II) starting from I by the alternate chemical oxidation. We assumed that if I carries 17-C00H function in place of 17-CHz as in cleanolic acid (IV), it would be appropriate to expect participation of the carboxylic function at the stage of intermediate (as in V) resulting 11a, 12a-epoxy-oleanolic lactone (VI) as a final product. On the other hand, a unique nortriterpenoid named eupteleogenin $(VII)^{2,3}$, a sapogenin of the biological active saponin, is so far the only natural occurrence possessing an 11α , 12α -epoxy-13, 28-lactone moiety, and as a model experiment for the biogenesis of the moiety, it seemed worthwhile to synthesize the epoxy-lactone system in one step according to the above mentioned assumption, which has now been realized although the yield of VI was rather low. It is a purpose of the present communication to describe the results obtained by photochemical oxidation of oleanolic acid.

On irradiation⁴⁾ of oleanolic acid (IV) in 95% ethanol (adjusted to PH 2.0 by HCl) for 80 hours at room temperature with slow stream of oxygen, a mixture* was obtained. After repeated silica gel column and thin layer chromatographic separations, the mixture afforded three products tentatively designated 0-1 (3%)**, 0-2 (trace), 0-3 (12.5%)** with the starting compound recovered (ca. 15%).

The major product, 0-3 (VIII), C₃₀H₄₈O₄(M⁺: 472), mp. 274-278°; IR (KBr, cm⁻¹): 3530, 1741; NMR***(τ): seven methyls, 6.84 (1H, t-like, characteristic to >C₍₃₎<u>H</u>-OH(eq.)), 6.20 (1H, broad

509

^{*} TLC of the total mixture disclosed the formation of small amount of oleanolic lactone (VI, without epoxy function), which was, however, not isolated in a pure state. ** Based on the pure compound obtained. *** Measured at 100 Mc. in CDCL₂.

510

singlet, $W_{\frac{1}{2}} = 7 \text{ cps}, C_{(12)} \underline{H} - OH(ax.))$, was assumed to be 12*a*-hydroxy-oleanolic lactone (VIII) by its physical properties, and the assumption was verified by comparison of its 3*β*-0-acetate (IX) with the authentic specimen prepared by Barton's procedure⁵.

The very minor product, <u>0-2</u>, mp. 251-255°; IR (KBr, cm^{-1}): 3510, no carbonyl absorption band, showed a molecular ion at m/e 442. However, its structure has not yet been clarified due to the shortage of the material.

The other product, <u>0-1</u> (VI), $C_{30}^{H} _{46}O_4$, mp. 269-271.5°, $(\alpha)_D^{23}$ + 20.7° (c, 1.0 in $CHCl_3$); IR (KBr, cm^{-1}): 3536 (OH), 1771 (¥-lactone) 870 (a significant absorption band appeared also in $VII^{(2)}$); Mass (m/e): 470 (M⁺), 189 (a most abundant peak above m/e 100); NMR (τ): seven methyls, 6.82 (1H, t-like, assigned to $> C_{(3)}$ H-OH(eq.)), 7.05* (2H, singlet, W_2^h =3 cps.), seemed to have an epoxy-lactone moiety based on its IR and NMR spectra. Thus, a singlet (2H) appearing at τ 7.05 quite resembles to the corresponding signal (τ 7.08, 2H, singlet²), W_2^h =3 cps.**) of eupteleogenin (VII)² ascribable to 11 β -H. Although the singlet due to the epoxidic protons of either VI or VII seems to be rather unusual comparing to the report by Tori et al⁷. (e.g., the epoxide protons of methyl 11 α , 12 α -epoxy-5 β -cholanate gave an AB quartet pattern with J=4 cps.) and to J value (4.5 cps.) of the epoxide protons of II¹. it would be approved by assuming unexpected equivalency of 11 β -H and 12 β -H in both VI and VII probably caused by the lactone ring***.

Finally, the structure VI for the compound <u>0-1</u> was proved by the following conversion. Thus, H_2SO_4 -MeOH treatment of VI furnished a keto-lactone (X), mp. above 300°. IR (KBr, cm⁻¹): 3560, 3430 (OH), 1770, 1703 (C=0), similarly as in the case of eupteleogenin²⁾, and the keto-lactone (X), C₃₀ $H_{46}O_4$ was unambiguously prepared by CrO₃ oxidation of IX followed by alkaline hydrolysis and relactonization****.

To shed further light on the participation of the other oxygen function at C_{17} , an analogous study on erythrodiol having a carbinol function at C_{17} (I, R=CH₂OH) is in progress in our laboratory.

^{*} The epoxidic protons are known to appear in the range $z 6.5-7.2^{6}$,

^{**} Dr. T.Murata of Res. Lab. of Takeda Chemical Industries very kindly allowed us to compare our spectra with his spectra of eupteleogenin derivatives², to whom we wish to express our sincere thanks. *** The signal due to 12β -H of II appeared at higher chemical shift (τ 7.21) than 11β -H (τ 6.88)¹, while 12β -H of VI or VII seems to be deshielded by the oxygen function nearby which may result the equivalency of 12β -H and 11β -H.

while $1\beta_{1}$ and $12\beta_{1}$ and $11\beta_{1}$. **** LiAlB₄ reduction of X yielded two tetrahydroxy derivatives, i.e., $3\beta_{1}, 2\beta_{1}$, $13\beta_{1}, 2\beta_{2}$ -tetrahydroxyand $3\beta_{1}, 12\alpha_{1}, 13\beta_{1}, 2\beta_{2}$ -tetrahydroxy-oleananes (XI,XII). An attempt (in a TLC scale) was made to prepare XII from VI by the epoxide ring opening with LiAlB₄ as reported in the case of eupteleogenin (VII)²), however the product was found non-identical with either XI or XII. Although we have no definite evidence at this stage of discussion, we assume that the epoxide ring of VI could be cleaved by LiAlB₄ leaving a hydroxyl function at 11α as in the case of II¹ rather than 12α as assumed in the paper of VII².



IV R=COOH oleanolic acid





VII eupteleogenin



IX R=Ac

We wish to express our deepest thanks to Res. Lab. of Takeda Chemical Industries for measuring NMR and mass spectra, and also to Drs. K.Morita and T.Murata of the Res. Lab. for the fruitful discussions.

References

- 1) I.Agata, E.J.Corey, A.G.Hortmann, J.Klein, S.Proskow, J.J.Ursprung: J.Org.Chem., 30 1698 (1965).
- 2) T.Murata, S.Imai, M.Imanishi, M.Goto, K.Morita: Tetrahedron Letters, 1965 3215.
- 3) M.Nishikawa, K.Kamiya, T.Murata, Y.Tomiie, I.Nitta: ibid., 1965 3223.
- 4) As the light source, a 100W high-pressure mercury lamp (Osawa Denki Co., Tokyo) was used. The similar result was obtained on using a 30W low pressure mercury lamp (Type PIL 30, Eiko-sha, Osaka).
- 5) D.H.R.Barton, N.J.Holness: J.Chem.Soc., 1952 78.
- 6) N.S.Bhacca, D.H.Williams, "<u>Applications of NMR spectroscopy in Organic Chemistry</u>", p.99 (Holden-Day, Inc., 1964, San Francisco).
- 7) K.Tori, T.Komeno, T.Nakagawa: J.Org.Chem., 29 1136 (1964).